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Development of a scalable synthesis of a nonbasic inhibitor of the serine protease tryptase

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Abstract—A chromatography-free process for the synthesis of a bis(benzimidazole)difluoromethane inhibitor of the serine protease tryptase is described. This synthesis features the introduction of the *gem*-difluoro moiety using the electrophilic fluorinating reagent N-fluoro-bis(phenylsulfonimide) as well as the stepwise introduction of both benzimidazole rings. A protocol for the destruction of reactive, process-related substances produced in the synthesis is also presented. © 2006 Elsevier Ltd. All rights reserved.

1. Introduction

Inhibition of mast cell tryptase has been recognized as a viable therapeutic target for allergic and inflammatory diseases such as allergic rhinitis, conjunctivitis, dermati-tis, inflammatory bowel disease (IBD), and asthma.^{[1](#page-3-0)} For example, a Phase IIa clinical evaluation of the tryptase inhibitor APC-366 (1) showed significance in the primary endpoint of reduction of the late asthmatic re-sponse when administered by inhalation.^{[2](#page-3-0)} In addition, in a Phase II clinical trial with patients exhibiting mildly to moderately active ulcerative colitis, intravenous administration of APC-2059 (2) resulted in 58% of these patients experiencing a clinically significant benefit in symptoms associated with the disease.^{[3](#page-3-0)} Other tryptase inhibitors, either in pre-clinical or clinical development, include RWJ-56423 (3) and BMS-354326 (4). Structures for these compounds are shown in [Figure 1](#page-1-0).

Another class of tryptase inhibitors is based on the 2,2'bis(benzimidazole)methane scaffold, including the symmetrical bis(5-amidino-2-benzimidazolyl)methane (BA-BIM; 5). The BABIM series afforded one of the first potent inhibitors of mast cell tryptase, having been initially reported as a potent inhibitor of other serine proteases in the late 1970s.[4,5](#page-3-0)

Research at Celera (formerly Axys Pharmaceuticals) led to the discovery of a novel binding mode of these inhibitors and determined that the potency of these compounds against trypsin-like proteases such as Factor Xa, and tryptase result from the tetrahedral binding of zinc(II) ions to both the hydroxyl group of Ser-195 and the imidazole moiety of His-57 in the active site of the enzyme, along with two nitrogens of the bis(benz-imidazole)methane moiety of the inhibitor.^{[6](#page-4-0)} This novel mechanism takes advantage of ambient levels of zinc in vivo, especially in the granules of the mast cell. However, development of orally active BABIM derivatives as serine protease inhibitors, including tryptase, were hampered by the presence of the polar benzamidine and metabolically labile benzylamine P_1 moieties, illustrated in the BABIM class (5) as well as the unsymmetrical derivative, APD-10 (6). In order to identify an orally active tryptase inhibitor based on the bis(benzimidazole)methane motif, efforts were undertaken to identify related derivatives which did not rely on the benzamidine or similar polar functionality for binding affinity in the P_1 pocket of the enzyme. Ultimately, SAR efforts identified CRA-9249 (7) as an orally active tryptase inhibitor devoid of these polar functionalities at P_1 ([Fig. 2](#page-1-0)). In order to provide multigram and ultimately larger quantities of 7 for pre-clinical evaluation, we set out to develop a scalable process for this compound.

2. Results and discussion

The initial route to CRA-9249 was based on a stepwise, 'one-pot' coupling of 2,2-difluoromalonic acid (8) with

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Figure 1. Representative tryptase inhibitors.

Figure 2. Bis(benzimidazole)methane derived tryptase inhibitors.

two different 1,2-diaminobenzene derivatives 9 and 11 to each carboxylate functionality of 8 with the expensive coupling agent bromotripyrrolidinophosphonium hexafluorophosphate (PyBroP) to give the bis(amide) $12^{7,8}$ $12^{7,8}$ $12^{7,8}$ Without isolation, compound 12 was heated in acetic acid to generate both benzimidazole rings of CRA-9249. Although this approach was satisfactory in providing modest amounts of 7 needed for early discovery efforts, the route suffers from low yield (20% from diacid 8 after column chromatography) and therefore was unsuitable for the manufacture of multigram to kilogram quantities of the active pharmaceutical ingredient (API, compound 7).

In order to arrive at a scalable process to 7, which circumvented the liabilities evident in the route described in Scheme 1, we pursued a stepwise condensation route

to the bis(benzimidazole)methane fragment. Generation of the gem-difluoromethane bridgehead would rely on the reaction of the bridgehead methylene of the bis benzimidazole moiety with an electrophilic fluorinating reagent. This transformation is based upon a successful strategy previously employed at Celera on a similar bis(benzimidazole)methane derivative.^{[9](#page-4-0)} The synthesis of the key bis(benzimidazole)difluoromethane acid 20 is shown in [Scheme 2](#page-2-0). Both benzimidazole rings in 20 were constructed from two separate 1,2-diaminobenzene derivatives according to this scheme. First, the righthand benzimidazole ring was constructed starting from

Scheme 1. Reagents and conditions: (a) PyBrOP, N-methylmorpholine (NMM), DMF, -10 °C to rt; (b) PyBrOP, DMF, -10 °C to rt; (c) 11, NMM, -10 °C to rt; (d) AcOH, 85 °C, 3 h.

Scheme 2. Reagents and conditions: (a) 40% aq CH₃NH₂, 100–105 °C; (b) 1 N aq HCl, 75% for both steps; (c) H₂, 5% Pd–C, THF–MeOH (8:3), 96%; (d) H₂, 10% Pd–C, EtOH, 89%; (e) EtOH, rt to 35–40 °C; (f) 15, DMPU, 150 °C, 90%; (g) (PhSO₂)₂N–F, 1,4-dioxane, reflux; 91%; (h) MeOH, reflux.

commercially available 3-methoxy-4-nitrobenzoic acid (13) which was converted to the 3-methylamino derivative 14 via a nucleophilic aromatic displacement of the methoxy group with 40% aqueous methylamine. Reduction of the nitro group in 14 with hydrogen and 5% palladium on carbon provided 4-amino-3-methylamino-benzoic acid 15.[7](#page-4-0)

The 3,5-difluoro-1,2-diaminobenzene required for the left-hand benzimidazole ring was produced by catalytic reduction of commercially available 3,5-difluoro-6-nitroaniline (16) with 10% Pd–C to afford the 3,5-difluoro-1,2-diaminobenzene (11). Reaction of the diamine 11 with the imidate of ethyl cyanoacetate $(17)^{10}$ $(17)^{10}$ $(17)^{10}$ in ethanol at 35–40 C provided the left-hand benzimidazole fragment 18 in 65% yield. Finally, thermal condensation of ester 18 with a degassed suspension of 3-methylamino-4-methyl benzoic acid 15 in DMPU at 150° C afforded the bis(benzimidazole)methanecarboxylic acid 19. [11](#page-4-0) Fluorination of this acid with an excess (250 mol $\%$) of *N*-fluorobenzenesulfonimide afforded the desired difluoro bridgehead intermediate 20 in 74% yield after heating the crude product in refluxing methanol. The hot methanol treatment served to degrade small amounts of the monofluoro- and keto-derivatives 21 and 22 derived from incomplete fluorination and competitive air oxidation of the methylene group in 19. Although we had determined that bis(benzimidazole)methyl derivatives similar to 19 were toxic, we suspected that the keto derivative 22, as well as the corresponding amide related to 7, could also be toxic. Furthermore, the chemical reactivity of 22 to hot methanol also supported our concerns that these ketones could also be toxic.

Hence, stringent specifications (<0.05%) for their presence were put in place for the final API. The treatment of crude 20 in refluxing methanol served to degrade the reactive monofluoro- and keto-derivatives 21 and 22 and reduce their levels below these stringent specifications prior to the introduction of the $2'$ -(4-fluorophenoxy)ethylamine fragment. The resulting degradants 23 and 24 were very soluble and easily removed during the methanol treatment.

Several routes were evaluated to prepare 4-fluorophenoxyethylamine derivative 29 required for the synthesis of CRA-9249 (7), which are described in [Scheme 3.](#page-3-0) The first route started from the Mitsunobu reaction of commercially available *N*-(tert-butoxycarbonyl)-aminoethanol 25 and 4-fluorophenol (26) using diisopropylazodicarboxylate (DIAD) and triphenylphosphine to provide Boc-protected 2-phenoxyethylamine 27. Unfortunately this approach was ultimately abandoned due the formation of modest amounts (15–20% as determined by proton NMR analysis) of the N-Boc aziridine (28), which required a tedious chromatographic separation to remove this potentially troublesome impurity from the desired N -Boc-aminophenoxyether.^{[12,13](#page-4-0)}

Although this route could be employed to afford the target amine hydrochloride 29 after deprotection with a solution of anhydrous HCl, a more scalable approach was developed, based on the Delepine reaction.^{[14](#page-4-0)} The

Scheme 3. Preparation of the right-hand amine 29. Reagents and conditions: (a) Ph₃P, THF, rt to $0-5$ °C; (b) *i*-PrO₂CN=NCO₂Pr-*i*, 0– 5 °C, then 0–5 °C to rt; (c) silica gel chromatography then 4 N HCl in 1,4-dioxane, CH₂Cl₂, 0–5 °C to rt (60% from 25); (d) CHCl₃, reflux, 95%; (e) EtOH, 12 N HCl; (f) 4 N aq NaOH, H2O, 88% for two steps; (g) anhyd HCl in 1,4-dioxane, then $Et₂O$, 85%.

commercially available 4-fluorophenoxy-ethylbromide (30) was reacted with hexamethylenetetramine (31) to provide the ammonium salt 32. Hydrolysis of 32 to the free amine followed by conversion to the HCl salt provided amine 29 on a multigram scale without chromatography. Several other routes were explored but none were as convenient as the Delepine reaction for large-scale preparations of amine 29.^{[15](#page-4-0)}

The coupling of amine 29 and acid 20 using mixed anhydrides afforded very good to excellent yields after addition of water to crystallize the product from N,Ndimethylformamide (DMF). We noted that the nature of the chloroformate employed in the reaction made a significant difference in conversion and yield of the crude product. Use of either isobutyl chloroformate $(R = i-)$ Bu) or isopropyl chloroformate $(R = i-Pr)$ afforded CRA-9249 (7) in excellent purity. However isopropyl chloroformate was found to be superior to isobutyl chloroformate in the conversion of 20 to 7. The latter reagent provided as much as 18% of unreacted 20 which fortunately could be easily recovered through acidification of the filtrate. Attempts to convert unreacted acid 20 through charging additional chloroformate to the reaction mixture resulted in the formation of side-products, such as compound 33 which is produced by N-acylation of the free N–H of the left-hand benzimidazole nitrogen and carbamate 34 which arises from competitive reaction of excess chloroformate with amine 30. Thus, 1 M

Scheme 4. Reagents and conditions: (a) DMF, NMM, $0-5$ °C; (b) ROCOCl, $(R = i-Pr \text{ or } i-Bu)$, $\leq 10 \degree C$; (c) 29, DMF; (d) NMM, $\leq 10 \degree C$ to rt; (e) recrystallization from ethanol.

isopropyl chloroformate in toluene was the preferred reagent to generate the mixed anhydride, ensuring >95% conversion and excellent yields of CRA-9249 (Scheme 4).

CRA-9249 (7) was isolated in 72% yield after chloroformate-mediated amide bond formation and recrystallization of the crude solid. Ultimately, this process was used to generate over 95 g of 7 in >99.0% purity. The limit of detection for the keto- and monofluoro impurities was less than 0.03% as determined by spiking experiments with authentic samples. The only impurity identified at significant levels $(>0.1\%)$ was the acid precursor 20 (0.25%) ^{[16](#page-4-0)}

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References and notes

- 1. (a) Bachelet, I.; Munitz, A.; Levi-Schaffer, F. Expert Rev. Clin. Immunol. 2005, 1, 63–73; (b) Cairns, J. A. Pulm. Pharmacol. Ther. 2005, 18, 55–66; (c) Burgess, L. E. Drug News Perspect. 2000, 13, 147–157; (d) Gangloff, A. R. Curr. Opin. Invest. Drugs 2000, 1, 79–85.
- 2. Krishna, M. T.; Chauhan, A. J.; Little, L.; Sampson, K.; Mant, T. G.; Hawksworth, R.; Djukanovic, R.; Lee, T. H.; Holgate, S. J. Allergy Clin. Immunol. 2001, 1077, 1039–1045.
- 3. Tremaine, W. J.; Brzezinski, A.; Katz, J. A.; Wolf, D. C.; Fleming, T. J.; Mordenti, J.; Stenkoski-Nix, L. C.; Kurth, M. C. Alim. Pharmacol. Ther. 2002, 16, 407–413.
- 4. Caughey, G. H.; Raymond, W. W.; Bacci, E.; Lombardy, R. J.; Tidwell, R. R. J. Pharmacol. Exp. Ther. 1993, 264, 676–682.
- 5. Tidwell, R. R.; Geratz, J. D.; Dann, O.; Volz, G.; Zeh, D.; Loewe, H. J. Med. Chem. 1978, 7, 613–623.
- 6. (a) Janc, J. W.; Clark, J. M.; Warne, R. L.; Elrod, K. C.; Katz, B. A.; Moore, W. R. Biochemistry 2000, 39, 4792– 4800; (b) Katz, B. A.; Clark, J. M.; Finer-Moore, J. S.; Jenkins, T. E.; Johnson, C. R.; Ross, M. J.; Luong, C.; Moore, W. R.; Stroud, R. M. Nature 1998, 391, 608–612.
- 7. Mittendorf, J.; Henning, R.; Raddatz, S.; Schlemmer, K.-H.; Hiraoka, M.; Kadono, H.; Mogi, M.; Moriwaki, T.; Murata, T.; Sakakibara, S.; Shimada, M.; Yoshida, N.; Yoshino. T. PCT Int. Appl. WO 0020401, 2000. CRA-9249 was developed by Bayer as part of an agreement between Bayer and Arris Pharmaceutical Corporation, now part of Celera. Celera re-acquired the tryptase program from Bayer in 2002.
- 8. Although bis(amide) 12 was not isolated, this intermediate could be produced as a mixture of regioisomers. The specific regioisomer shown here is based on the assumption that acylation occurs on the more basic nitrogens found on each 1,2-diaminobenzene derivative 9 and 11.
- 9. Sperandio, D.; Gangloff, A. R.; Litvak, J.; Goldsmith, R.; Hataye, J. M.; Wang, V. R.; Shelton, E. J.; Elrod, K.; Janc, J. W.; Clark, J. M.; Rice, K.; Weinheimer, S.; Yeung, K.-S.; Meanwell, N. A.; Hernandez, D.; Staab, A. J.; Venables, B. L.; Spencer, J. S. Bioorg. Med. Chem. Lett. 2002, 12, 3129–3133.
- 10. Imidate 17 was prepared in 85% yield on a 1.1 mol scale from the reaction of ethyl cyanoacetate and HCl gas in a mixture of toluene and ethanol $(\sim 6:1)$ at 0–5 °C.
- 11. The conditions used to produce 19 also afforded small amounts of the decarboxylated bis(benzimidazole)methane derivative shown below, whose identity was confirmed by comparison to an authentic sample.

- 12. The preparation of compound 29 by alkylation of α chloroacetamide with 4-fluorophenol followed by $LiAlH₄$ reduction has been described in the literature: Shtacher, G.; Taub, W. J. Med. Chem. 1966, 9, 197–203. However, we did not evaluate this process for the work described here.
- 13. Protection of commercially available 2-bromoethylamine hydrobromide with di-tert-butyl dicarboxylate and Nmethylmorpholine (NMM), followed by bromide displacement with 4-fluorophenol and cesium carbonate also provided 27 contaminated with aziridine 28.
- 14. (a) Galat, A.; Elion, G. J. Am. Chem. Soc. 1939, 61, 3585– 3586. For a review of synthetic applications of hexamethylenetetramine, including the Delépine reaction, please see the following: (b) Blažević, N.; Kolbah, D.; Belin, B.; Šunjić, V.; Kajfež, F. Synthesis 1979, 161–176.
- 15. Other routes explored in the preparation of 29 included reduction of nitrile 35 and reaction of bromide 30 with ammonia. Reaction of 35 using either catalytic [e.g., H_2 / PtO2; transformation (a)] or chemical reduction [e.g., BH_3 THF or LiAlH₄, THF, reflux; transformation (b)] were unsatisfactory, providing either C–O bond cleavage as the primary reaction pathway or formation of small amounts of N, N -bis[2-(4'-fluorophenoxy)ethyl]amine (36). The hydrogenolysis of the C–O bond of α -phenoxyacetonitriles is a known process: Benarab, A.; Boyé, S.; Savelon, L.; Guillaumet, G. Tetrahedron Lett. 1993, 34, 7567–7568, Displacement of the bromide in 30 with ammonia in several solvent combinations and conditions (transformation c) was also plagued by formation of 36 and the corresponding trialkylamine which could not be easily separated from the desired product.

16. The 95-g lot of 7 also provided acceptable analysis results including combustion (CHN), inductively coupled plasma (ICP), residue on ignition (ROI) and moisture content (Karl Fischer).

LC-MS: mlz 516.0 [M+1]⁺; ¹H NMR (DMSO- d_6 ; 400 MHz) δ 8.8 (t, 1H), 8.3 (s, 1H), 7.85 (d, 1H), 7.8 (d, 1H), 7.3 (d, 1H), 7.2 (t, 1H), 7.1 (m, 2H), 7.0 (m, 2H), 4.1 (t, 2H), 4.0 (s, 3H), 3.7 (q, 2H); CHN: calcd: C, 58.26; H, 3.52; N, 13.59; F, 18.43. Found: C, 58.03; H, 3.28; N, 13.62; F, 18.55; ICP results: Zn: <1 ppm; Cu < 1 ppm. ROI: <0.10%; % moisture (Karl Fischer): <0.10%.